Electro-Organic Synthesis of New Pyrimidine and Uracil Derivatives

Saied Saeed Hosseiny Davarani,* Neda Sheijooni Fumani, Siavash Vahidi, Mohammad-Ali Tabatabaei, and Hamid Arvin-Nezhad

Department of Chemistry, Faculty of Science, Shahid Beheshti University, G.C. Tehran 1983963113, Iran *E-mail: ss-hosseiny@cc.sbu.ac.ir Received December 30, 2008 DOI 10.1002/jhet.231 Published online 21 December 2009 in Wiley InterScience (www.interscience.wiley.com).



Electrochemical oxidations of catechols have been studied in the presence of 6-amino pyrimidine derivatives as nucleophiles in aqueous solution using cyclic voltammetry. The efficient electro-organic synthesis of products has been successfully performed at carbon rod electrodes in an undivided cell under controlled potential conditions in good yield and purity.

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INTRODUCTION

Pyrimidine and its derivatives are attracting the attention of an increasing number of synthetic organic chemists because of their broad range of biological activity and medicinal importance [1,2]. Numerous reports delineate the antitumor [3], antiviral [4], antioxidant [5], antifungal [6], and hepatoprotective [7], activities of these compounds. Therefore, large efforts for the preparation of these molecules have been directed toward the synthetic manipulation of uracils [8]. The importance of pyrimidine derivatives prompted us to synthesis a number of these compounds from catechols and thiouracils. We have investigated the electrochemical oxidation of catechols (1a-1c) in the presence of 6-amino pyrimidine derivatives (3a, 3b) as nucleophiles. This work has led to the development of a facile and environmentally friendly electrochemical method for synthesis of pyrimidine derivatives (**5a–5c**) and uracil derivatives (**8d–8f**).

RESULTS AND DISCUSSION

Cyclic voltammetry of 2 m*M* solution of catechol (**1a**) in 0.2*M* sodium acetate solution containing 10% acetonitrile as supporting electrolyte shows an anodic (A₁) and a corresponding cathodic peak (C₁), which correspond to the transformation of (**1a**) to *o*-quinone (**2a**) and *vice versa* within a quasi-reversible two electrons reaction (Fig. 1, curve a). A peak current ratio (I_P^{C1}/I_P^{A1}) of nearly unity can be considered as a criterion for the stability of *o*-quinones (**2a**) produced at the surface of the electrode under the experimental conditions [9,10]. In other words, any hydroxylation [11,12], dimerization





Figure 1. Cyclic voltammogram of 2 m*M* catechol (**1a**): (a) in absence of 6-amino-1-methyl-2-(methyl)2-(methylthio)pyrimidin-4(1*H*)-one (**3a**), (b) in the presence of 6-amino-1-methyl-2-(methyl)2-(methylthio)pyrimidin-4(1*H*)-one, (c) cyclic voltammogram of 2 m*M* 6-amino-1-methyl-2-(methyl)2-(methylthio)pyrimidin-4(1*H*)-one (**3a**) in the absence of catechol, at glassy carbon electrode, in 0.2*M* sodium acetate solution containing 10% acetonitrile. Scan rate: 100 mVs⁻¹, *T* = ambient temperature. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

[13,14], or radical cations formation [15–18] are too slow to be observed on the time scale of cyclic voltammetry.

To get further support on the electrochemical oxidation of catechol (1a), it was studied in the presence of 3a as a nucleophile. Curve b in Figure 1 shows the cyclic voltammogram obtained for a 2.0 mM solution of 1a in the presence of 2.0 mM 3a. The voltammogram exhibits decreasing in cathodic counterpart (C₁) of anodic peak (A₁). This is due to the reactivity of 1a with 3a. The cyclic voltammogram of 2.0 mM of 3a is shown in Figure 1 curve c, for comparison.

The multicyclic voltammogram of **1a** in the presence of 6-amino-1-methyl-2-(methyl)2-(methylthio)pyrimidin-4(1H)-one (**3a**) are shown in Figure 2. In this figure, the second scan exhibits a relatively intense decrease in anodic peak current A₁ together with a potential shift in a positive direction. The decrease in A₁ peak current and positive shift of this peak are probably due to the formation of a thin film of product at the surface of the electrode inhibiting to a certain extent the performance of electrode process [19,20].

Furthermore, it was observed that the height of the C_1 peak increased proportional to augmentation of potential scan rate (Fig. 3, curve a–f). This confirms the reactivity of **2a** toward **3a**. A similar situation was observed when

Figure 2. Cyclic voltammogram of 2 m*M* catechol: (a) in the presence of 6-amino-1-methyl-2-(methyl)2-(methylthio)pyrimidin-4(1*H*)-one (first cycle), (b) in the presence of 2 m*M* 6-amino-1-methyl-2-(methyl)2-(methylthio)pyrimidin-4(1*H*)-one (second cycle), at glassy carbon electrode, in 0.2*M* sodium acetate solution containing 10% acetonitrile. Scan rate: 100 mV s⁻¹, T = ambient temperature. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

3a/1a concentration ratio is decreases. Moreover, the current function for A₁ peak $(I_P^{A1}/v^{1/2})$ decreases slightly with increasing scan rate (Fig. 3, curve g).

Controlled-potential coulometry was performed in an aqueous solution containing 0.5 mmol of **1a** and 0.5 mmol of **3a** at the potential of A_1 peak. At the end of coulometry, it was specified that charge consumption per molecules of **1a** becomes about $2e^-$.

The coulometry and voltammetry results allow us to propose an EC mechanism[16,17] for the electro-oxidation of 1a in the presence of 3a (Scheme 1). According to our results, the Michael addition reaction of 3a to oquinone (2a) [eq. (2)] seems to occur much faster than other side reactions, which leads to the product 5a. The overoxidation of 5a was circumvented during the preparative reaction because of the almost insolubility of the product in acetate buffer solution medium.

The electro-organic synthesis of **5b** and **5c** has been performed using oxidation of **1b** and **1c** in the presence of **3a** as described for **5a** (Table 1).

The electro-oxidation of catechol (1a) in the presence of 6-amino-2,3-dihydro-2-thioxopyrimidin-4(1*H*)-one (3b) as a nucleophile was studied by cyclic voltammetry and controlled-potential coulometry in 0.2M sodium acetate solution containing 10% acetonitrile too. Figure 4, curve b, shows the cyclic voltammogram obtained for a 2.0 mM



Figure 3. Typical cyclic voltammogram of 2 m*M* catechol (1a) in the presence of 2 m*M* 6-amino-1-methyl-2-(methyl)2-(methylthio)pyrimidin-4(1*H*)-one (3a) in 0.2*M* sodium acetate solution containing 10% acetonitrile at a glassy carbon electrode (1.8-mm diameter) at various scan rate. Scan rate from (a) to (f) are 50, 100, 200, 400, 800, and 1600 mV s⁻¹, respectively. (g) Variation of peak current ratio $(I_P^{A1}/v^{1/2})$ versus scan rate, T = ambient temperature. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

solution of **1a** in the presence of 2.0 m*M* **3b**. The voltammogram clearly exhibits an increase in anodic peak A_1 and a decrease in the cathodic peak C_1 . This is due to the reactivity of **2a** with **3b**. For comparison, the cyclic voltammogram of 2.0 m*M* solutions of catechol (**1a**) and **3b** are shown in Figure 4, curves a and c, respectively.

The possible reason for the observed large increase in the A_1 peak current could be the oxidation of





Figure 4. Cyclic voltammogram of 2 m*M* catechol (**1a**): (a) in the absence of 6-amino-2,3-dihydro-2-thioxopyrimidin-4(1*H*)-one (**3b**), (b) in the presence of **3b**, (c) cyclic voltammogram of 2 m*M* 6-amino-2,3-dihydro-2-thioxopyrimidin-4(1H)-one (**3b**) in the absence of catechol, at glassy carbon electrode, in 0.2*M* sodium acetate solution containing 10% acetonitrile. Scan rate: 100 mV s⁻¹, *T* = ambient temperature. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]



intermediate **5c** at a potential close to that of the starting catechol. This oxidation is due to solubility of the **5c** in the reaction medium. The most important differences between this and previous cases are the large increase in the A_1 peak current (Fig. 4, curve b) and the number of transferred electrons during controlled-potential coulometry.

The results demonstrate that contrary to the previous cases, the consumed charge is about $4e^-$ per molecule of catechol (1a). This is related to two two-electron transfer processes [eqs. (1) and (4) in Scheme 2]. The coulometry, voltammetry, and NMR results allow us to propose an ECEC mechanism [21–24], indicated in

Scheme 2 for the electro-oxidation of catechol (1a) in the presence of 3b.

CONCLUSIONS

In conclusion, the results of this work show that catechols (1a-1c) are oxidized to their respective o-benzoquinone (2a-2c). The formed o-benzoquinones are attacked by nucleophiles 3a and 3b to form final products 5a-5c and 8d-8f. We observed an interesting diversity in the electro-oxidation mechanism and products of catechols (1a-1c) in the presence of (3a, 3b). In the cases of 1a-1c in the presence of 3a, the final products **5a–5c** are pyrimidine derivatives that were obtained after consumption of 2e⁻ per molecule 1a, 1b, and 1c. In the case of 1a-1c in the presence of 3b, the final products 8d-8f are uracil derivatives that were obtained after consumption of 4e⁻ per molecule **1a–1c**, *via* intermolecular and intramolecular Michael addition reactions. The overall mechanism for anodic oxidation of catechols (1a-1c) in the presence of 3a and 3b are presented in Schemes 1 and 2. These mechanisms show a good diversity in anodic oxidation of 1a-1c in the presence of 3a and 3b.

EXPERIMENTAL

Chemical and solutions. Catechol derivatives were reagentgrade materials, sodium acetate, and other solvents were of proanalysis grade (all from E. Merck). These chemicals were used without further purification. The stock solution of catechols was prepared daily.

Electrode and electrochemical instrument. A cyclic voltammetry was performed using a micro-Autolab type III potentiostat/galvanostat and millimole scale electrolysis was performed using a pp-200 Zahrner potentiostat/galvanostat. The working electrode used in voltammetry experiments was a glassy carbon disc (2.7 mm^2 area) and a platinum wire was used as the counter electrode. The working electrode used in controlled-potential coulometry and millimole scale electrolysis was assembly of three carbon rods (27 cm^2 area) and large platinum gauze constituted the counter electrode. The working electrode potentials were measured *versus* 3M Ag/AgCl reference electrode (carbon rods from Azar electrode and other electrodes from Metrohm).

Bruker IFS-66 FTIR spectrometer, Shimadzu QP 1100-EX mass spectrometer operating at an ionization potential of 70 eV, and Bruker DRX-300 AVANCE NMR spectrometer were used for recording different spectra.

Electrochemical synthesis of 5a–c and 8d–f. In a typical procedure, 100 mL of 0.2*M* sodium acetate solution containing 10% acetonitrile was pre-electrolyzed at mentioned potential (Table 1) *versus 3M* Ag/AgCl in an undivided cell. Then 2 mmol of catechols (1a–1c) and 2 mmol of nucleophiles (3a, 3b) were added to the cell. Initially, the current density was \sim 2 mA/cm² and the electrolysis was terminated when the decay of the current became more than 95%. The process was

 Table 1

 Electro-analytical and preparative data.

Conversion	Applied potential (V) vs. (Ag/AgCl)	Yield (%)	Consumed charge (C)	Time of electrolysis (h)
1a–5a	0.3	68	98.1	16.3
1b–5b	0.3	72	97.8	16.2
1c-5c	0.3	69	98.2	16.4
1a-8d	0.25	64	196.2	33.4
1b-8e	0.25	67	196.8	33.1
1c8f	0.25	62	196.5	33.8

interrupted during the electrolysis and the graphite anode was washed in acetone to reactivate it. At the end of electrolysis, a few drops of acetic acid were added to the solution and the cell was placed in a refrigerator overnight. The precipitated solid was collected by filtration and purified by washing with hot water (Table 1).

Characteristics of products

6-Amino-5-(3,4-dihydroxyphenyl)-3-methyl-2-(methylthio)pyrimidin-4(3H)-one (5a). Mp > 270°C; IR (KBr) (ν_{max} cm⁻¹): 3438, 3324, 3225, 2931, 1635, 1590, 1537, 1505, 1418, 1359, 1223, 1092, 845. ¹H NMR (DMSO-d₆): δ = 3.30 (s, 3H, CH₃), 3.36 (s, 3H, CH₃), 5.86 (s, 2H, NH₂), 6.31 (d, J = 8Hz,1H, Ar–H), 6.45 (s, 1H, Ar–H), 6.59 (d, J = 10Hz, 1H, Ar–H), 8.16 (s, 1H, OH), 8.79 (s, 1H, OH) ppm; ¹³C NMR (DMSO-d₆): δ = 14.6, 29.9, 95.1, 106.1, 111.7, 124.2, 133.3, 146.0, 148.6, 158.0, 159.8, 160.8 ppm; MS, m/z (%): 279 (M⁺, 100), 230 (10), 171 (40), 126 (30), 110 (50), 83 (46), 63 (50), 47 (90). Anal. Calcd. for C₁₂H₁₃N₃O₃S: C, 51.60; H, 4.69; N, 15.04. Found: C, 51.48; H, 4.73; N, 14.98.

6-Amino-5-(3,4-dihydroxy-5-methylphenyl)-3-methyl-2-(methylthio)pyrimidin-4(3H)-one (5b). Mp > 270°C; IR (KBr) (v_{max} cm⁻¹): 3445, 3157, 2930, 1636, 1516, 1411, 1359, 1298, 1204, 1093, 1036, 906, 860. ¹H NMR (DMSO-d₆): δ = 2.09 (s, 3H, CH₃), 2.41 (s, 3H, CH₃), 3.16 (s, 3H, CH₃), 5.78 (s, 2H, NH₂), 6.42 (s, 1H, H-Ar), 6.53 (s, 1H, H-Ar), 8.11 (s, 1H, OH), 9.09 (s, 1H, OH) ppm. ¹³C NMR (DMSO-d₆): δ = 14.6, 16.5, 29.9, 95.1, 115.7, 123.5, 124.3, 124.5, 142.4, 145.0, 158.0, 159.7, 160.9 ppm; MS (EI, 70 eV): m/z (%) = 293 (M⁺, 100), 243 (5), 171 (25), 124 (50), 105 (20), 88(50), 57 (55), 41 (50). Anal. Calcd for C₁₃H₁₅N₃O₃S: C, 53.23 H, 5.15; N, 14.31. Found: C, 53.18; H, 5.18; N, 14.31.

6-Amino-5-(3,4-dihydroxy-5-methoxyphenyl)-3-methyl-2-(methylthio)pyrimidin-4(3H)-one (5c). Mp > 270°C; IR (KBr) (v_{max} cm⁻¹): 3456, 1636, 1583, 1523, 1388, 1251, 1041, 998, 844, 800, 578. ¹H NMR (DMSO- d_6): $\delta = 3.21$ (s, 3H, OCH₃), 3.49 (s, 3H, CH₃), 3.71 (s, 3H, CH₃) 5.86 (s, 2H, NH₂), 6.33 (s, 1H, H-Ar), 6.62 (s, 1H, H-Ar), 8.35 (s, 1H, OH), 9.85 (s, 1H, OH) ppm.¹³C NMR (DMSO- d_6): $\delta = 19.3$, 23.4, 29.9, 90.2, 105.7, 121.4, 125.2, 145.4, 153.2, 157.2, 159.9, 161.0, 163.4 ppm; MS (EI, 70 eV): m/z (%) = 309 (M⁺, 100), 236 (10), 171 (12), 140 (32), 88(25), 57 (30), 41 (50). Anal. Calcd for C₁₃H₁₅N₃O₄S: C, 50.47 H, 4.89; N, 13.58. Found: C, 50.46; H, 4.89; N, 13.60.

2,3-Dihydro-6,7-dihydroxy-2-thioxo-1H-pyrimido[4,5-b]indol-4(9H)-one (8d). Mp > 270°C; IR (KBr) (v_{max} cm⁻¹): 3415, 3343, 3173, 1639, 1566, 1481, 1450, 1366, 1309, 1201, 864. ¹H NMR (DMSO-*d*₆): $\delta = 5.03$ (s,1H, NH), 639 (s, 1H, H-Ar), 6.70 (s, 1H, H-Ar), 7.20 (s, 1H, NH), 8.38 (s, 1H, NH), 9.37 (s, 1H, OH), 9.41 (s, 1H, OH) ppm; ¹³C NMR (DMSO*d*₆): $\delta = 81.1$, 106.4, 108.6, 112.3, 129.3, 144.9, 145.1, 161.0, 162.4, 163.0 ppm; MS (EI, 70 eV): m/z (%) = 249 (M⁺, 80), 221 (25), 210 (25), 182(90), 155(10), 128 (10), 85 (30), 68 (100), 41 (55). *Anal.* Calcd for C₁₀H₇N₃O₃ S: C, 48.19; H, 2.83; N, 19.25. Found: C, 48.22; H, 2.85; N, 19.75.

2,3-Dihydro-6,7-dihydroxy-8-methyl-2-thioxo-1H-pyrimido[**4,5-b**]indol-**4(9H)-one** (**8e**). Mp > 270°C; IR (KBr) (v_{max} cm⁻¹): 3464, 3367, 2928, 1633, 1505 1452, 1361, 1297, 1228, 1033, 804. ¹H NMR (DMSO-*d*₆) δ = 2.17 (s, 3H, CH₃), 5.01 (s, 1H, NH), 6.94 (s, 1H, H-Ar), 7.28 (s, 1H, NH), 8.22 (s, 1H, NH), 8.86 (s, 1H, OH), 9.97 (s, 1H, OH) ppm; ¹³C NMR (DMSO-*d*₆) δ = 14.6, 81.2, 105.7, 113.9, 117.3, 118.0, 128.2, 142.6, 144.9, 161.1, 162.4 ppm; MS (EI, 70 eV): m/z (%) = 263 (M⁺, 100), 223 (15), 196 (75), 150 (10), 124 (25), 85 (30), 68 (60), 41 (100). *Anal.* Calcd for C₁₁H₉N₃O₃ S: C, 50.18; H, 3.44; N, 15.96. Found: C, 50.17; H, 3.45; N, 15.97.

2,3-Dihydro-6,7-dihydroxy-8-methoxy-2-thioxo-1H-pyri*mido*[**4,5-b**]*indo*1-**4**(**9H**)-*one* (**8***f*). Mp > 270°C; IR (KBr) (v_{max} cm⁻¹): 3435, 3336, 3339, 2924, 1659, 1565, 1463, 1394, 1278, 1020, 768. ¹H NMR (DMSO-*d*₆) δ = 2.93 (s, 3H, OCH₃), 4.91 (s, 1H, NH), 6.11 (s, 1H, H-Ar), 6.68 (s, 1H, NH), 7.88 (s, 1H, NH), 8.35 (s, 1H, OH), 10.31 (s, 1H, OH) pm; ¹³C NMR (DMSO-*d*₆) δ = 59.6, 91.5, 108.2, 121.4, 128.9, 132.2, 150.9, 152.6, 158.5, 166.3, 170.1 ppm; MS (EI, 70 eV): m/z (%) = 279 (M⁺, 93), 251 (70), 223 (15), 157 (50), 140 (30), 110 (27), 60 (80), 41 (100). *Anal.* Calcd for C₁₁H₉N₃O₄S: C, 47.31; H, 3.25; N, 15.05. Found: C, 47.29; H, 3.26; N, 15.06

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